ION BEAM METHODS FOR THE SURFACE CHARACTERIZATION OF POLYMERS

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<u>Abstract</u> - Ion beam methods of surface characterization have not been applied extensively to polymers and other organic materials, but would appear to be very useful for these materials. Applications of high energy ion beam methods would seem to be limited except for specific cases but low energy methods such as ion scattering spectrometry (ISS) and secondary ion mass spectrometry (SIMS) should have numerous uses with polymeric materials. ISS is a true surface method which determines elemental composition at the first monolayer, but tells little about how the elements are combined. SIMS has high sensitivity for many elements, and has the capability of determining something of the molecular structure near the surface. Both ISS and SIMS, as well as high energy methods, give elemental composition with depth. These surface spectroscopies are useful in many areas of polymer technology including synthesis, extrusion and forming, and long time durability and stability under thermal and electromagnic radiation.

# INTRODUCTION

Surface molecular structure and elemental composition of polymers have been largely ignored, even though the number of surface sensitive techniques has increased rapidly during the past few years. There are probably two main reasons that surface analysis methods have been applied extensively to metals and alloys but only to a limited degree to polymers and other organic materials. Most surface analysis methods are carried out in very clean ultrahigh vacuum systems which may be contaminated by organic materials. Even just the <u>possibility</u> of contamination usually keeps the organic material <u>outside</u> the vacuum system. The other main obstacle to surface characterization of organic materials is that they are usually very good insulators. When these materials are bombarded with ions, electrons or x-rays, the usual surface probes, they build up a charge which can confuse the interpretation of data or even make it impossible to obtain spectra. There are various compensation methods which will be described, but charging still remains a problem.

Previously surface analysis of polymers has been limited essentially to the use of x-ray photoelectron spectrometry (XPS) (Ref. 1,2). Such an example is the study of the effect of an oxygen plasma treatment on the adhesion of metal films on polymers (Ref. 3). These studies provide elemental and chemical combination data obtained from core level photoemission spectra. However, Gardella and Hercules (Ref. 4), have pointed out some inherent limitations of the technique with respect to its application to most polymer systems. Further, they suggest that photoelectron escape depth is sufficient to cause averaging from several atom layers, approaching bulk results. For these reasons it is of interest to explore alternative methods of surface analysis for polymers. Surface analysis techniques based on probing the specimen with ions provide new methods for analysis of polymers. Some of the most promising of these techniques will be described here.

## PRINCIPLES AND PROCEDURES

## Ion Scattering Spectrometry (ISS)

Use of low energy backscattered ions to characterize a surface is a relatively recent development. The method has been reviewed by Buck (Ref. 5). High energy ions had been used in the past to analyze surfaces but it was not until Smith (Ref. 6) used low energy (1 KeV) noble gas ions to probe the surface of a variety of materials that the technique came into popular use. It was found from this work and others that when the energy of ions was lowered the scattered ion spectra became simpler and sharper and event from a single surface atom. Therefore, the energy  $E_1$  retained by an ion of mass,  $M_{\rm ion}$  with an incident energy  $E_0$  after

PAAC54:2 F

scattering from an atom of mass  $M_{atom}$  through an angle  $\theta$  is given by equation (1) which is based on the conservation of kinetic energy and momentum (here  $M_{ion}$  is smaller than  $M_{atom}$ ).

$$\frac{E_1}{E_0} = \frac{M_{ion}^2}{(M_{ion} + M_{atom})^2} \qquad \left\{ \cos\theta + \left[ \frac{M_{atom}^2}{M_{ion}^2} - \sin^2\theta \right]^{\frac{1}{2}} \right\}^{\frac{1}{2}}$$
(1)

For 90° scattering which is frequently used this reduces to a very simple relationship:

$$E_1/E_0 = (M_{atom} - M_{ion})/(M_{atom} + M_{ion}).$$
 (2)

The experimental setup for 90° scattering instruments is shown in Figure 1A and is representative of original commercial instrumentation (3M Company, St. Paul, Minnesota) which used a 127° electrostatic analyzer. A great improvement in sensitivity was gained by the development of a cylindrical mirror analyzer (CMA) substituted for the original electrostatic sector as shown in Fig. 1B.

Fig. 1. Experimental set-up for Ion Scattering Spectrometry



The geometry of the CMA results in a scattering angle of 138°. The major advantage of low energy ion scattering is the extremely fine surface selectivity when low energy ions collide with the surface atom. The probability for neutralization is very high because of the long residence time ( $10^{-14}$  to  $10^{-16}$  sec.). Only about one in  $10^3$  of the scattered particles retain a positive charge even after one collision. Therefore, the probability that an ion is still in the charged state after two or more collisions is very small. Since the detector responds only to charged particles, contributions from particles which scatter more than once are almost certainly neutral and not counted by an ion detector. These facts suggest that an instrument using time-of-flight methods to detect either ions or neutrals would be extremely useful. An inherent feature of ion scattering which may be considered an advantage is the simultaneous sputtering of the surface as energy is transferred to the surface atoms from the ion beam. It is an advantage in that the concentration of the various atomic species may be followed with depth. On the other hand it is a disadvantage because damage is being produced by the sputtering. Once the atom sputters from the surface the sample is changed, and an exact experiment on that spot may not be repeated. However, ion current density may be kept very low by rastering the beam over a large area, thus minimizing surface damage. One positive feature of ion scattering compared to most other spectroscopies is the simplicity of the spectra. Binary ion scattering results in one peak for each isotope of an element present. For instance the scattering of helium from teflon in Fig. 2 results in the appearance of only two peaks in the spectrum, one for carbon and one for fluorine. Each peak is very sensitive to the amount present but absolute quantitative analyses can be difficult since the scattered yield depends on scattering cross section and neutralization efficiency, neither of which is well known for most elements. During the ion scattering experiment atoms are sputtered from the surface, allowing depth profiling analysis from the removal of the surface layers by the probe ion. Use of helium ions gives a very slow rate of surface removal. Neon and argon provide much higher sputtering rates or the ion beam may be focused and rastered on the surface to reduce sputtering while the signal is gated from the center of the crater to minimize

crater edge effects. The signal may be collected and imaged from the surface using the rastered beam to give a lateral analysis of the surface. Therefore ion scattering provides a combination of in depth analyses and lateral analyses to give a three dimensional picture of the chemical makeup of the surface.

Fig. 2. Ion Scattering Spectrum using  ${}^{3}\text{He}^{+}$  for teflom



Secondary Ion Mass Spectrometry (SIMS)

When a low energy ion strikes the solid surface it undergoes a number of interactions which are outlined in Table 1 (Ref. 7) and illustrated in Fig. 3.

TABLE 1. Effect of ion impact on a solid surface including emission processes & changes in the surface zone

EMISSION PROCESSES

CHANGES IN THE SURFACE OF THE TARGET

Atomic and molecular particles Neutrals Positive ions Negative ions Excited particles Loss of surface particles Sputtering Recoil implantation

Implantation Primary ions Surface atoms (recoil)

Electrons Surface processes (Auger de-excitations, e.g.) Bulk processes (ionization, e.g.)

Photons

Gas phase processes Surface processes Bulk processes Imperfections Amorphization Chemical effects

Lattice destruction

Breaking of bonds Bond formation

The process under consideration here is process #5 as seen in this figure (the reflected ion giving energy to the surface atom which is sputtered). The sputtered species which are removed from the surface are made up of both positive and negative ions as well as neutral particles. Neutral particles have much more abundance than ionic species and have also been used for surface analysis. Surface analysis by SIMS falls into two categories, low current density sputtering and high current density sputtering. Categories are determined by the characteristics of the primary ion beam. A low current density sputtering analysis results in a very small fraction of the surface being disturbed, a result that approaches a basic requirement of a true surface analysis method. This is generally known as the Static SIMS (SSIMS) method. High current density sputtering removes more material and is required for preparing elemental depth profiles. In the high current density method, changes are seen in the surface and near surface regions. Table 2 shows the main features of SIMS as a surface analysis method (Ref. 7). Of the positive attributes listed probably the extremely high sensitivity for many elements is the greatest advantage of SIMS. On the other hand, the large differences in sensitivity for different surface structures and chemical combination is the largest negative factor involved in SIMS analyses. Isotopic identification and the sensitivity to hydrogen are two other important uses of the SIMS method. These advantages and disadvantages are reviewed by Werner (Ref. 9) and quantitative aspects are discussed by the same author (Ref. 10).



TABLE 2.	Main	features	of	SIMS	as	а	surface	analysis	method	(Ref.	7)

Positive	<ul> <li>Information depth in the "monolayer range"</li> <li>Detection of all elements including hydrogen</li> <li>Detection of chemical compounds</li> <li>"Lateral resolution" in the range of atomic distances</li> <li>Isotope separation</li> <li>Extremely high sensitivity for many elements and compounds (&lt;10<sup>-6</sup> monolayers)</li> <li>Quantitative analysis after calibration</li> <li>Negligible destruction of the surface (SSIMS)</li> <li>Elemental profiling (Dynamic SIMS)</li> </ul>
Negative	<ul> <li>Large differences in sensitivity for different "surface structures" (factor 1000)</li> <li>Problems in quantitative interpretation of molecular spectra</li> </ul>

- Ion induced surface reactions

Equipment for SIMS may be as simple as that shown in Fig. 4 or as complex as the ion microprobe. In a simple system a SIMS experiment required a vacuum chamber to house the experiment, a sample holder, an ion source, an energy analyzer and a mass analyzer. In such simple systems the noble or reactive gas fills the system and the entire chamber including the ion gun and sample area are at approximately  $1-5 \times 10^{-5}$  torr.

Fig. 4. Simple block diagram showing components of a SIMS system



A more complicated type of instrument seen in Fig. 5 is one in which the performance is improved through the use of a differentially pumped vacuum system to produce ultra high vacuum in the vicinity of the sample.

> =⊗== Argon entry Primary ion source 10 cm Quadrupole gas Wien filter analyzer electromagnet 阆 Reactive gas entry Secondary ion quadrupole mass filte Automátic pressure controller Ш 11 ( Viewport łίc Г Electron multiplier Ionization Electron guage aun

This also allows the entry of a reactive gas in the sample chamber area while sputtering with a noble ion for studying chemical changes or reactions of the surface. Still another improvement and added complexity may be made to the SIMS instrument by the mass analysis of the primary beam. The energy filter is generally made up of several elements whose function is to optimize collection of the secondary ions and to filter and focus the ions at the entrance to the mass analyzer. The mass analyzer in simple systems is usually a quadrupole filter. Colton and Coworkers (Ref. 12) show equipment using a quadrupole filter capable of detecting mass species at more than 5000 amu. It is recognized that SIMS has been used successfully as a stand-alone technique to solve and to perform surface physics research (see review by Colton, Ref. 13). However, it appears that the area of greatest use of SIMS thus far is as a complement to other surface characterization methods. The extremely high sensitivity for some elements can be taken advantage of by using SIMS with other techniques in which these elements do not show high sensitivity. The SIMS technique is also ideal to use with fundamentally low resolution methods such as ion scattering, to separate and identify the adjacent masses which may be present at the sample surface. The most popular combination of instru-ments used thus far has been ISS-SIMS and AES-SIMS. SIMS has also been used on scanning electron microscopes, allowing high quality imaging along with lateral and depth analysis of the sample.

## OTHER TECHNIQUES USING AN ION BEAM

#### Rutherford Backscattering

When a surface is bombarded with a beam of ions in the MeV energy range a small fraction of the incoming particles undergoes a Rutherford collision and is backscattered. A recent book has been published on this subject (Ref. 14). The experiment is conceptually simple but requires complicated and expensive equipment to carry it out. Typical experimental set-up for routine backscattering analysis is shown in Fig. 6 (Ref. 14). Charged particles are generated in an ion source, the energy is then raised to several MeV by an accelerator, usually a van de Graaff. The high energy beam then passes through a series of devices which will focus the beam and filter it for a particular type of particle and energy. The beam then enters the scattering chamber and impinges on the sample. Some of the backscattered particles strike the detector where they generate an electrical signal. This signal is amplified, processed with analog and digital electronics. The energy E of the ion scattered through an angle of 180° is related to the initial energy E<sub>0</sub> by the equation:

$$E = \left[\frac{M-m}{M+m}\right]^2 E_0$$
(3)

where m is the mass of the scattered particle and M is the mass of the target atom. Thus energy analysis of the primary ions backscattered from a specimen provides a mass analysis over a sample volume with depth. The relative sensitivity of different surface atoms is determined by the Rutherford cross-section for the element. There is also an effect on the energy of the sample thickness by the ion beam and so therefore the energy spectrum is a

# Fig. 5. Diagram of equipment for SIMS (Ref. 11)

convolution of a mass scale established by the backscattering process and a depth scale established by an energy loss of the ion beam before and after the backscattering event. Probably there is little application for this technique to polymers, except perhaps for coatings, metallized plastics, or to determine homogeneity in filled plastics.

Fig. 6. Experimental arrangement for high energy ion backscattering analysis (Ref. 14)



<u>Surface composition by analysis of neutral ion impact radiation (SCANIIR)</u> In addition to causing sputtering at the surface, a primary ion beam causes the emission of photons in the visible and ultraviolet region (Ref. 15). By analyzing the light output with an optical spectrometer the chemical analysis of the surface may be obtained and depth profiling obtained. SCANIIR has not come into general use but would appear to be a useful technique for surface analysis especially as a complementary technique with some of the other methods.

#### INFORMATION OBTAINED BY THESE TECHNIQUES

The two ion beam techniques most popularly used for true surface analysis are Ion Scattering Spectrometry and Secondary Ion Mass Spectrometry. Both of these techniques have certain capabilities and limitations. These features are summarized in Table 3 (Ref. 13).

# TABLE 3. Comparison of ion spectroscopies

Parameter	Ion Scattering Spectrometry (ISS)	Secondary Ion Mass Spectrometry (SIMS)
Principle	elastic binary collision with surface ion	sputtering of surface atoms by ion beam
Probe	~1 to 3 keV ions	~1 to 3 keV ions
Signal	ion current versus energy	ion current versus mass
Applicable elements	Z <u>&gt;</u> 3	all (if positive and negative SIMS)
Surface sensitivity	high	variable
Elemental profiling	yes	yes
Image-spatial analysis	yes	yes
Spectral shift	possible, but generally no	no
Information on chemical combinations	yes, in fine features but generally no, expecially pol	in some cases (fingerprint spectra)

Quantitative analysis	yes	maybe with similar standards
Influence of operating conditions and matrix	no	yes
Isotopic analysis	yes, in principle but generally no because or resolution limits	yes
Beam induced surface changes	yes, sputtering damage (minimal with low fluence)	yes, sputtering damage (minimal in "static" mode)

These two techniques may be applied in a gentle manner as described earlier using a near static ion beam to produce little change in the surface and also in a mode in which chemical profiling with depth is possible. ISS can detect all of the elements heavier than helium in the periodic table. The sensitivity variation across the periodic table is probably less than one order of magnitude. SIMS provides a distinct advantage of being able to analyze, in principle, all of the elements in the periodic table. Being able to identify isotopes is a definite advantage. The sensitivity of the SIMS technique can vary several orders of magnitude (perhaps up to 10<sup>4</sup>) due to a rapidly changing secondary ion yield caused by matrix and chemical effects. The ability of ion scattering to resolve different elements in a complete unknown is at times somewhat limited. There are few intrinsic limitations or spectral interferences but the technique is fundamentally a low resolution technique in which there may be some uncertainty as to the exact identity of a given line. Specificity may be improved by going to a scattering ion closer to the mass of the unknown element, that is, we would use helium for the light elements usually found in polymers, neon for intermediate mass elements, and argon for the heavy elements.

Ion scattering gives very little information on the chemical combination of the element detected in the sample, however, recently discovered yield variations and the use of other fine features in the spectrum give some possibility of using ion scattering to determine chemical species at the surface but probably not in polymers. The appearance of cluster ions in the secondary ion mass spectrum gives a good possibility of using SIMS to determine chemical combinations and deduce molecular structure in polymers. The interpretation of such spectra is extremely complicated and has to be treated with a great deal of care. Molecular ions can be dislodged from the surface and give some idea of the chemical combination, but molecular complexes may also be synthesized at the sample surface in the gas phase above the sample surface. The presence of such ions in the mass spectrum does not unequivocally prove the presence of such species in or on the sample itself. However, a detailed analysis allows identification of fragments split from a parent molecule, and often the parent mass or "m + 1" peak is seen in the spectrum. Sufficient resolution in the mass spectrum is often required to separate species occurring at the same nominal mass number. Some of these interferring species are shown in Table 4 (Ref. 17 from unpublished data of Evans).

TABLE 4.	Types of	interferences	and	typical	examples	(Ref.	17	)
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Interference	Interfering	Analyt.	Required
type	ion	ion	resolution
Multiply charged	28 Si2+	14N+	950
matrix ion	62Ni2+	31Р⁺	3200
Matrix selfpolymers	<sup>16</sup> 0, <sup>+</sup>	32S+	1800
ions	28Si2+	56Fe+	2950
Prim. ion-matrix	Cu <sub>3</sub> 0+	207Pb +	1050
molecular ions	Si <sub>2</sub> 0+	75As +	3250
	AlO <sub>2</sub> +	59Co +	1500
Hydride ions	<sup>30</sup> SiH <sup>+</sup>	31 <b>p +</b>	4000
	FeH +	55Mn +	3300
	SnH+	121 Sb +	19500
Hydrocarbon ions	C.H.+	27A1 +	650
	C H	63Cu +	650
	C <sub>2</sub> H <sub>2</sub>	CN	2000

#### SPECIMEN CHARGING

A serious problem encountered in ion beam methods of analysis is that of specimen charging. Impact of energetic positive ions causes development of a positive charge on the surface of an insulator. This effect is especially prevalent in polymers, because they are usually excellent insulators. Table 5 lists work functions for some typical polymers. The high value of these work functions shows the reason for the charging exhibited by polymers. TABLE 5. Work function of polymers (Ref. 18)

Polymer	Work Function, $\phi$			
Teflon Chlortrifluoroethylene Vinyl Chloride Sulfone Styrene Methyl Methacrylate	5.75 5.30 5.13 4.95 4.90 4.68			
Nylon 6.6	4.30			

This charge will influence or even prevent the emission of secondary ions. To overcome this charging of insulators there have been numerous methods used, as seen in Table 6.

TABLE 6. Methods of change neutralization (Ref. 17)

- (1) Deposition of a conducting thin film or of a grid.
- (2) Use of Cs<sup>+</sup> as primary ions, in this way a conducting layer is continuously built up by the primary ion bombardment.
- (3) Compensation of the charging by means of an extra electron beam.
- (4) Use of neutral beams: the charging is reduced from the case of positive primary ions.
- (5) Application of special electrodes for draining excessive negative charge.
- (6) Shift of target holder potential  $V_{\rm H}$  by  $\Delta V$  in a direction opposite to the previous charging of the insulator.

Some of these have significant limitations or contaminate the surface by coating a conductor on the surface of the insulator. In the mechanism of the charging of the insulator surface the impact of the positive ions cause secondary electron emission and consequently positive charge buildup. The obvious way of removing this charge is then to flood the surface with a beam of low energy electrons restoring charge neutrality. This method of charge compensation by electron bombardment is shown from work of Muller (Ref. 19) in Fig. 7. Muller's method has the advantage that deflecting the electron beam means that the surface of the sample cannot see the hot electron source and therefore cannot be contaminated by material boiled from the electron source. Since there is no direct line of sight, there is little heating caused by electrons from the filament. An example of SIMS spectra in the high mass range when using the charge compensation method of Muller, is seen in Fig. 8. Here strong, sharp symmetrical peaks are observed well out into the mass 300 range from the polymer Teflon, which has a very high work function as shown in Table 5. Muller's method uses very low energy thermal electrons (>5eV). Wittmaack (Ref. 20) used a focussed beam of intermediate energy electrons (100-500eV), while Magee and Harrington (Ref. 21) used well defined electrons in the energy range 500-2000eV for charge compensation. Whatever the exact method used, charge neutralization is necessary for most polymers to obtain either SIMS or ISS data. The effects which may be caused by neutralization include contamination, heating and diffusion or transport of mobile ions. These potential problems should be considered when analyzing insulators.

Fig. 7. Charge compensation in a SIMS instrument (Ref. 19)



Fig. 8. SIMS data in mass range 160-330 from Teflon using charge neutralization (Ref. 19)



Mass spectrum (linear scale) for the positive secondary ions from Teflon up to m/e = 330

## APPLICATIONS

As mentioned earlier, ion beam methods have not been applied extensively to organic materials and polymers. Ion scattering has been applied to adhesive bonding materials to determine the locus of failure (Ref. 22) and contamination effects (Ref. 23). DiBennedetto and Scola (Ref. 24) have used both ISS and SIMS to characterize surfaces of treated glass fibers and fiber/ polymer interfaces. The results show how SIMS can be used to study the chemical surface and chemical changes on the surface and at interfaces. By working at low power levels with insulator surfaces, the SIMS analysis showed changes in the structure of a polymerized silane coating as a function of depth of penetration into the interface. The concentration of nitrogen and hydrogen generated from the surface maintained a relatively constant level as the distance from the airsilane interface increased; then within 160 A into the surface, a dramatic increase in the nitrogen level was noted, to a depth of 240A. In this region, the simplicity of the SIMS spectra, with major peaks corresponding to the atomic constituents of  $\mathbf{y}$ -aminopropyltriethoxysilane, namely, H, C, N, O, and Si, suggested that low-molecularweight oligomer was present in this region. This means that the silane coating was not sufficiently cured to provide a mechanically stable interface. Finally, from 240 A to the silane-glass interface, the nitrogen and hydrogen generated from the surface reached a lower constant level but about three times higher than that generated from the air-silane domain. This suggests that the silane polymer coating adjacent to the glass interface is different from the silane polymer at the air interface. Thus, it is clear that the ISS/SIMS technique can be used to define the interface and interphase regions and also to follow changes at the interface due to a chemical reaction.

Gardella and Hercules (Ref. 14) have shown STMS data for poly (alkyl methacrylates) and also shown ISS data for Teflon. They found changes in molecular fragmentation patterns with only very slight changes in polymer processing. Careful examination of core level XPS data shows no identifiable changes in core level binding energies or intensity ratios. Side chain structure in the ester portion of the poly (alkyl methacrylates) dramatically influenced static SIMS data.

There have been no papers published concerned only with ISS of polymers except for abstracts of meeting presentations by Sparrow and Mishmash in 1977 (Ref. 25) and Gardella and Hercules in 1979 (Ref. 26). Both ISS and SIMS data appear in papers in which the emphasis is in another area, such as the cleaning of surfaces by ultraviolet light (Ref. 27). Figure 9 shows ISS/SIMS data from that work for polypropylene and polypropylene exposed to UV light. Although the spectral changes are subtle they are typical of changes seen during long time aging and weathering. Usually, following UV exposure, slightly more oxygen is observed in the ISS data. SIMS data show some changes in the fragmentation pattern and larger fragments are observed. Perhaps further study of such changes may be attributed to bond breakage and cross linking effects.

Fig. 9 (a) ISS/SIMS data for polypropylene using <sup>3</sup>He<sup>+</sup> at 2500 eV.
(b) ISS/SIMS data for polypropylene exposed to low pressure mercury discharge lamp, same conditions as in (a).



Even on similar linear polymers, SIMS shows a different fragmentation pattern as can be seen in Fig. 10, ISS/SIMS data for polyethylene.

Fig. 10. ISS/SIMS data for polyethylene, using  $^{3}\text{He}^{+}$  at 2500 eV.



On the other hand, the ion scattering spectrum is very <u>non</u> individualistic. In fact, the ISS data for graphite in Fig. 11, is so similar to spectra obtained from straight chain polymers, that they probably could not be distinguished from each other.

Fig. 11. ISS/SIMS data for graphite using <sup>3</sup>He<sup>+</sup> at 2500 eV.



SIMS, on the other hand, shows fragmentation pattern changes even on the same material but given different treatment. Figure 12 shows SIMS data for a commercial two part epoxy mixed under the same conditions and then divided into two portions, one cured 24 hours at room temperature and the other cured one hour at 250°F.

Fig. 12. Secondary Ion Mass Spectra from commercial two part epoxy cured at room temperature and at 250°F.



As can be seen, some larger fragments are seen in the sample held at elevated temperature, and sodium has segregated to the surface. Such segregation is very common in high temperature cured specimens, where sodium is often found at the failure surface in an adhesive failure mode. ISS/SIMS data from the adhesive side of a titanium-epoxy failure interface from a tensile test specimen are shown in Fig. 13.

Fig. 13. ISS/SIMS data from adhesive side of failure surface from adhesive bonded test specimen



The fragmentation pattern is different from this temperature sensitive tape epoxy and sodium is seen at the failure interface. Sodium was also observed on the matching titanium side of the specimen.

Although little data for polymers has yet made its way to the literature, SIMS has been applied to other solid organic materials. Karasek (Ref. 28) in 1974 showed that parent masses and predictable fragments were observed in the + SIMS data for organic materials such as benzoic acid and chlorobenzoic acid, even when extremely energetic 8KeV argon ions were used to obtain spectra. Benninghoven and his associates have applied near static conditions to obtain spectra from a wide variety of materials ranging from amino acids, nucleic acids, peptides, drugs and vitamins (Refs. 29-31). An example of both positive and negative SIMS data from a peptide (Ref. 30) is seen in Fig. 14 (Ref. 30) and for vitamin C in Fig. 15 (Ref. 31). Note the  $(M+1)^+$  and  $(M-1)^-$  ions present in the spectra. Absolute yield of ions for a number of organic materials has been tabulated by Benninghoven and Sichtermann and are tabu-

lated in Table 7 (Ref. 31). Note the often high yield on decarboxylated species in amino acids.

Fig. 14. Positive and negative SIMS data for phenylalanylglycine on Ag using argon primary ion at 2.25 KeV.



Fig. 15. Positive and negative SIMS data for ascorbic acid (Vitamin C) on Ag (Ref. 31).



334

TABLE 7.	Absolute yield of "parent lil	e" secondary	ions of	organic	compounds	on
	silver (Ref. 31).					

			yield $S(X) \times 100 L$			
	formula	mol wt	(Number per i	of secondary incident prima	ions × 100, ry ion)	
I. amino acid			(M + H) <sup>+</sup>	(M – H)⁻	(M – COOH)*	
glycine	C <sub>2</sub> H,NO,	75	120.0	_	52.0	
α-alanine	C,H,NO,	89	21.0	40.0	53.0	
β-alanine	C <sub>3</sub> H <sub>2</sub> NO <sub>2</sub>	89	88.0	19.5	7.2	
phenylalanine	C,H <sub>11</sub> NO,	165	4.0	0.3	13.0	
serine	C <sub>3</sub> H,NO <sub>3</sub>	105	61.0	18.0	61.0	
threonine	C₄H,NO,	119	8.3	2.6	13.8	
proline	C,H,NO,	115	19.2	8.8	72.0	
valine	C,H,,NO,	117	8.0	8.3	32.0	
leucine	C <sub>6</sub> H <sub>1</sub> ,NO <sub>2</sub>	131	0.8	26.4	40.0	
norleucine	C,H,NO	131	24.8	6.5	76.0	
arginine	C <sub>6</sub> H <sub>14</sub> N <sub>4</sub> O <sub>2</sub>	174	7.2	2.4	2.1	
tyrosine	C,H, NO,	181	7.4	·	13.6	
tryptophan	$C_{11}H_{12}N_{2}O_{2}$	204	3.5	0.8	3.5	
cysteine	C'H'NO'S	121	12.0	11.0	15.0	
cystine	C, H, N, O, S,	240	4.0	1.6	1.8	
methionine	C,H <sub>11</sub> NO,S	149	13.1	5.4	9.4	
ethionine	C <sup>4</sup> <sup>1</sup> ,NO <sup>2</sup> S	163	13.6	5.6	12.0	
giutamine	C <sub>5</sub> H <sub>10</sub> N <sub>2</sub> O <sub>3</sub>	146	7.2	8.3	4.3	
II. derivates of amino acids			(M – H) <sup>-</sup>	$(M' - H)^{-a}$	(M – Cl)*	
glycine ethylester HCL	C <sub>4</sub> H <sub>10</sub> CINO <sub>2</sub>	139	-	1.6	180.0	
alanine ethylester HCL	C, H, CINO,	153	-	-	48.0	
cysteinium HCL	C,H,CINO,S	157	-	4.0	19.7 <sup>6</sup>	
taurine	C <sub>2</sub> H <sub>7</sub> NO <sub>3</sub> S	125	4.8	-	-	
III. peptides			(M + H)*	(M – H)⁻	(M – COOH)	
glycylglycine	C <sub>4</sub> H <sub>4</sub> N <sub>2</sub> O <sub>3</sub>	132	41.6	4.8	-	
glycylglycylglycine	C,H,N,O,	189	4.0	0.4	2.0	
glycylleucine	C,H,N,O,	188	1.6	4.2	3.0	
phenylalanylglycine	C <sub>11</sub> H <sub>14</sub> N <sub>2</sub> O <sub>3</sub>	222	8.0	1.6	-	
IV. drugs			(M + H) <sup>∗</sup>	(M – H) <sup>-</sup>	(M – OH)*	
barbital	C <sub>8</sub> H <sub>12</sub> N <sub>2</sub> O <sub>3</sub>	184	-	44.0	-	
ephedrine	C <sub>10</sub> H <sub>15</sub> NO	165	16.0	-	40.0	
atropine	C <sub>17</sub> H <sub>23</sub> NO <sub>3</sub>	289	84.8 <sup>c</sup>	-	-	
epinephrine	C,H,NO,	183	-	6.4	-	
V. vitamins			(M + H) <sup>∗</sup>	(M – H)⁻		
ascorbic acid (C)	C'H'O'	176	3.7	17.6		
biotin (H)	C10H16N2O3S	244	0.3	4.2		
nicotinic acid (PP)	C,H,NO,	123	-	46.4		
nicotinamide	C <sub>6</sub> H <sub>6</sub> N <sub>2</sub> O	122	2.1	$15.2^{d}$		
VI. sulfonamides			(M + H)*	(M – H) <sup>-</sup>		
sulfanilic acid	C,H,NO,S	173	-	16.3		
sulfanilamide	C,H,N,O,S	172	0.6	17.6		
sulfacetamide	C <sub>8</sub> H <sub>10</sub> N <sub>2</sub> O <sub>3</sub> S	214	-	20.8		
VII. other compounds			(M + H) <sup>*</sup>	(M − H) <sup>-</sup>		
thymidine	C <sub>10</sub> H <sub>14</sub> N <sub>2</sub> O <sub>5</sub>	242	1.9	1.3		
manie	C, H, CIN,	259	-	-	96.0°	
acriflavine	19 19 3				f	
acriflavine creatine	C,H,N,O,	131	2.9	-	3.4'	

# Potential Applications

There are many possible areas of polymer technology in which ion beam methods of analysis could be used. In the synthesis of polymers, solid monomers, catalysts, and co-catalysts could be analyzed by secondary ion mass spectrometry. In the extrusion or other forming process, the distribution of processing aids such as lubricants could be determined. During use or accelerated testing the polymer could be analyzed to determine stability to electromagnetic radiation. Some specific areas and potential materials in which surface analysis might prove beneficial are listed in Table 8.

TABLE 8. Potential materials applications of surface analysis methods.

- A. Adhesives epoxy-phenolic, polyimide, etc.
- B. Coatings polyimides, Teflon, etc.
- Composite structure polybenzimidizoles, polyimides, etc. with fiberglas с.
- D. Fibers - aromatic polyamides, etc.
- E. Films Polyimide (Kapton), polyteser (Mylar), metallized polymers, etc.

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