Versatility of β-lactams in synthesis. Studies directed toward the synthesis of complex nucleoside antibiotics and some macrocyclic peptides*

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Abstract: The diastereoselective [2+2] cycloaddition of α -hydroxyketene equivalents with chiral α,ω -oxyaldehyde-derived imines followed by the 2,2,6,6-tetramethylpiperidinyl-1-oxyl (TEMPO)-promoted ring expansion of the resulting α -hydroxy β -lactam adducts provides an unconventional and short route to α -amino acid *N*-carboxy anhydrides (NCAs). The required enantiopure α,ω -oxyaldehydes were obtained either from the chiral pool or through the Sharpless AD methodology. Following the present strategy, several nonproteinogenic NCAs were synthesized, which were further coupled with α -amino acid esters giving rise to key fragments of some nucleoside antibiotics and macrocyclic peptides.

INTRODUCTION

In recent years, the way β -lactams are viewed has changed considerably. Presently, their interest stems not only from being a key structural feature of β -lactam antibiotics, but also from their valuable utility as synthetic intermediates, mainly as masked α - and β -amino acid derivatives. In fact, the cleavage of the otherwise constrained β -lactam ring at either N1–C₄ or N1–C₂ bonds has opened the way to a wide array of α -amino acid and β -amino acid-derived structures [1]. On the other hand, the development of highly stereoselective methods of forming the substituted azetidin-2-one ring [2] has given additional power to such a β -lactam route. In this context, we have been involved in the study of the potential of β -lactams as intermediates in synthesis and have discovered (Scheme 1) that α -hydroxy β -lactams,



Scheme 1 General strategy for the access to NCAs through enantiopure β -lactam intermediates.

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upon treatment with nitroxide-free radicals, such as 2,2,6,6-tetramethylpiperidinyl-1-oxyl (TEMPO), in combination with a solution of commercial bleach, undergo an unprecedented ring expansion to give α -amino acid *N*-carboxy anhydrides (NCAs) [3]. The well-recognized importance of this particular class of mixed anhydrides for peptide coupling [4] led us to develop this approach into a general method for the synthesis of short peptide segments containing α -amino β -hydroxy and α -amino β , ω -polyhydroxy acids, the key components of complex nucleoside antibiotics and some macrocyclic peptides.

$\alpha\textsc{-}AMINO\ \beta\textsc{-}HYDROXY\ ACIDS.\ SYNTHETIC\ APPROACH TO THE NONPROTEINOGENIC AMINO\ ACIDS\ OF\ LYSOBACTIN$

The practicability of the above strategy relies primarily on the efficiency, in terms of both chemical yield and diastereoselectivity, of the [2+2] hydroxyketene-imine cycloaddition reaction [5]. In particular, the cycloaddition of hydroxyketenes with α -oxyaldehyde-derived imines fulfills these requirements, and we have successfully applied this methodology, in a sequential manner, to the synthesis of the key tripeptide fragment present in the macrocyclic antibiotic lysobactin I [3b,6] (Fig. 1).



Fig. 1 Chemical structure of lysobactin. The tripeptide segment approached is framed.

Thus, the stereoselectively formed β -lactams 2 (Scheme 2) were transformed into the NCA 3a and 3b. Compound 3a was first opened by *S*-leucine methyl ester, and the resulting dipeptide adduct was coupled with 3b to afford, after amine deprotection-reprotection steps, tripeptide 4.



Scheme 2 Synthesis of tripeptide 4 by sequential opening of NCAs 3a and 3b.

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SYNTHETIC APPROACH TO PEPTIDYL NUCLEOSIDE ANTIBIOTICS FROM CHIRAL POOL SOURCES

Several nucleoside antibiotics are structurally comprised of a polyhydroxylated α -amino acid fragment coupled to another α -amino acid bearing a nucleoside unit. In the case of polyoxins **5** (Fig. 2), the polyhydroxylated α -amino acid is identified as polyoxamic acid **6** (R=H), and much effort has been devoted to its synthesis [7].



Fig. 2 Structure of some polyoxins.

Our approach began from commercially available tartaric acid esters, which can be easily transformed into the Mukaiyama's aldehyde through procedures well established in the literature [8]. Imine 7 (Scheme 3), directly formed from such an aldehyde, was then subjected to [2+2] cycloaddition to afford β -lactam 8 as single stereoisomer. Further oxidative ring expansion promoted by TEMPO gave rise to NCA 9, which was smoothly coupled with the desired α -amino acid ester to yield polyoxamic acid-derived peptides 10 [9]. Following the same strategy, compounds 11 and 12 were also synthesized [10]. It is worth mentioning that the degree of epimerization during the coupling of 9 with amino acid esters proved solvent-sensitive. While in either CH₂Cl₂ or Et₂O the isomerization degree was below the limit of detection (less than 0.5%), it was significant in more polar solvents (MeCN, 15%, MeNO₂, 8%; DMF, 50%; HMPA, 72%).



Scheme 3 Synthesis of polyhydroxylated α -amino acid-containing peptides.

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SYNTHESIS OF THE SOUTHWEST TRIPEPTIDE SEGMENT OF ECHINOCANDIN B. AN ALTERNATIVE STRATEGY TO THE CHIRAL POOL APPROACH

Finally, the above strategy has been slightly modified, *vide infra*, to accomplish the synthesis of peptides incorporating β , γ -dihydroxy α -amino acids. These subunits are present, for example, in echinocandin B (Fig. 3), a cyclic hexapeptide with potent antifungal activity [11].



Fig 3 Chemical structures of several echinocandins isolated from fungi.

In this instance, the α , β -dihydroxy aldehydes required as starting materials did not come from the chiral pool [8]. Instead, they were prepared in a stereodivergent way by the Sharpless AD technique [12] (Scheme 4). The [2+2] cycloaddition of either acetoxy- or benzyloxyketene to the corresponding α , β -dihydroxy aldehyde-derived imines **15** resulted critical. In general, cycloadducts **16** were obtained as single stereoisomers, while poorer diastereomeric ratios (typically 75:25) were attained for adducts **17**.



Scheme 4 Sharpless AD, [2+2] oxyketene-imine cycloaddition and final ring expansion sequence giving rise to enantiopure NCA 19.

With NCA **19** in hand, the synthesis was concluded (Scheme 5) with the coupling of **19** with the serine derivative **20** to afford dipeptide **21**, which, after protecting group manipulation and further peptide coupling with the 4-hydroxyproline derivative **24**, gave rise to the protected tripeptide **25** in good yield [13].

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Scheme 5 Final coupling steps toward the southwest tripeptide segment of echinocandin B.

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